

# BUKTI KORESPONDENSI

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Judul Jurnal	:	The phytochemical screening and antioxidant potential of <i>Cinnamomum javanicum</i> Blume leaves from Central Kalimantan

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5	Acceptance of manuscript (LoA)	06 Maret 2021	16
6	Article Published	01 Juni 2021	17
7	Ethical approval	21 Juli 2020	34

## 1. Register Akun Jurnal

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## 2. Submission article

Manuscript ID: BFSA-2102-1065

Manuscript Title: **The Phytochemical Screening and Antioxidant Potential of Cinnamomum javanicum Blume Leaves from Central Kalimantan**

Authors: syahrida dian ardhany, Susi Novaryatiin, Nanang Hanafi

Dear **Ms. syahrida dian ardhany**

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It should be noted that the manuscript will be reviewed for possible publication in the Scientific Journals Management System.

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## 3. Review Process (Manuscript Needs Revision)

Manuscript ID: BFSA-2102-1065

Manuscript Title: **The Phytochemical Screening and Antioxidant Potential of Cinnamomum javanicum Blume Leaves from Central Kalimantan**

Authors: syahrida dian ardhany, Susi Novaryatiin, Nanang Hanafi

Dear **Ms. syahrida dian ardhany**

Evaluation process of the above mentioned manuscript has been reviewed. The comments of the reviewer(s) are included at the bottom of this letter.

The reviewer(s) have recommended publication, but also suggest some revisions to your manuscript. Therefore, I invite you to respond to the reviewer(s) comments and revise your manuscript within the period of defined time.

Because we are trying to facilitate timely publication of manuscripts submitted to journal, your revised manuscript should be uploaded as soon as possible. If it is not possible for you to submit your revision in a reasonable amount of time, we may have to consider your paper as a new submission.

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Reviewers Recommendation:

### Reviewer 1:

Reviewer Comment For Author:

The authors should take the following conditions before the acceptance of this work.

- 1- The paper should determine the exact weight of the fresh leaves as well as after extraction to calculate accurately the percentage of each contents.
- 2- Authors should uniform the abbreviation (mL) or (ml)
- 3- The authors should rewrite the antioxidant activity test with DPPH method to be more clear.

### Reviewer 2:

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Reviewer Comment For Author:

- Many typos mistake.
- The methods used may be reduced and referred only by the references , no need to write them in details as they are well known methods.
- The results and the discussion parts should be written in a detailed way. It's very shallow.

potential presence of alkaloid, flavonoids, tannins, saponins, and steroids by using the following standard methods.<sup>13-16</sup>

*Determination of the total alkaloid content*

~~10-Ten~~ mg extract of *C. javanicum* was weighed then dissolved in 10 ml ethanol ~~pro-analysis~~.

~~1-One~~ ml extract was measured the absorbance with a spectrophotometer at 272 nm. The standard used for the calibration curve ~~is-was~~ caffeine.<sup>17</sup> The total alkaloid content was expressed as micrograms of alkaloids per ~~miligramsmilligrams~~ of the extract.

*Determination of the total flavonoid content*

~~5-Five~~ mg extract of *C. javanicum* was weighed then dissolved in 10 ml ethanol ~~pro-analysis~~.

~~1-One~~ ml extract ~~put-was placed~~ in the volumetric flask, ~~add-1one~~ ml AlCl<sub>3</sub> (2% b/v) and 8 ml acetic acid (5% v/v) ~~were added~~. After mixing, the solution was incubated for 20 minutes.<sup>18-19</sup> The solution was measured the absorbance with a spectrophotometer at 412 nm.

The standard used for the calibration curve ~~is-was~~ quercetin.<sup>20</sup> The total flavonoid content was expressed as micrograms of flavonoids per ~~miligramsmilligrams~~ of the extract.

*Determination of the total tannin content*

A total of 30 mg of sample was weighed and put into a 10 mL volumetric flask. The solution

## Introduction

Central Kalimantan is one of the tropical forest areas in Indonesia which has biodiversity especially natural medicine but has not been fully utilized. Traditional uses such as traditional medicine have been done, but further exploration and development have not been done much. Sintok lancang leaves (*Cinnamomum javanicum* Blume.) are one of the typical Central Kalimantan plants that have not been studied much, it's found in the Mungku Baru forest (Kawasan Hutan Dengan Tujuan Khusus) or educational forest managed by the Muhammadiyah University of Palangkaraya collaboration with the Borneo Nature Foundation (BNF). Local people use sintok lancang leaves for various diseases such as to treat abdomen pain, wounds, and diabetes.

There have not been many studies on *Cinnamomum javanicum*, especially those that grow in Indonesia, but one of the researches at the University of Jember states that different species, *Cinnamomum Sintoc* leaves has the potential as an antioxidant because it contains secondary metabolites such as flavonoids, alkaloids, steroids, terpenoids, and tannins which could prevent the free radical.<sup>1-3</sup> Antioxidant use is associated with reduced production of ROS (Reactive oxygen species) and free radicals.<sup>4-6</sup> According to previous pieces of evidence, ROS have been implicated in the pathogenesis of various diseases such as cancer, aging, and other diseases. In addition, research related to natural antioxidants are considered to contribute to the therapeutic approach in the treatment of COVID-19, although their mechanism against COVID-19 is still unknown and speculative.<sup>7</sup> Several studies provide evidence that a reduction in ROS accumulation slows the apoptosis signaling activated by a coronavirus.<sup>8</sup>

In this study, the determination was carried out the total flavonoids, alkaloids and tannins compounds of *Cinnamomum javanicum* leaves. Antioxidant activity test was conducted by using DPPH and FRAP assay. The present data would help to explore the potential of Indonesia's natural plants, especially plants that have not been studied much so it can be further developed for the cosmetic, food and pharmaceutical industries.

## **Methods**

### *Plant Collection and Identification*

Fresh leaves of sintok lancang (Fig 1.) were collected in the Mungku Baru forest (Kawasan Hutan Dengan Tujuan Khusus) or educational forest managed by the Muhammadiyah University of Palangkaraya collaboration with the Borneo Nature Foundation (BNF), Palangka Raya, Central Kalimantan and authenticated by Dr. R. Hendrian, M.Sc from Indonesia Institute of Sciences, Research Center for Plant Conservation and Botanic Gardens, Bogor, Indonesia.



**Fig 1.** Sintok lancang (*Cinnamomum javanicum* Blume.)

#### *Preparation of Plant Extract*

The whole fresh leaves were washed, cleaned and dried in oven at 45<sup>0</sup>C.<sup>9-10</sup> Generally, the oven drying method is used for herb drying in the temperature range of 40-60<sup>0</sup>C.<sup>11</sup> The dried leaves were grinded and powdered then extracted with ethanol 96% by percolation. Percolation is better than maceration because it is a continuous process in which the saturated solvent is constantly being replaced by fresh solvent.<sup>12</sup> The extract were concentrated in a rotary evaporator.

#### *Phytochemical Qualitative Screening*

The ethanolic extract of sintok lancang leaves (*C. javanicum* Blume) was screened for potential presence of alkaloid, flavonoids, tannins, saponins, and steroids by using the following standard methods.<sup>13-16</sup>

#### *Determination of the total alkaloid content*

Ten mg extract of *C. javanicum* was weighed then dissolved in 10 ml ethanol. One ml extract was measured the absorbance with a spectrophotometer at 272 nm. The standard used for the calibration curve was caffeine.<sup>17</sup> The total alkaloid content was expressed as micrograms of alkaloids per milligrams of the extract.

#### *Determination of the total flavonoid content*

Five mg extract of *C. javanicum* was weighed then dissolved in 10 ml ethanol. One ml extract was placed in the volumetric flask, one ml AlCl<sub>3</sub> (2% b/v) and 8 ml acetic acid (5% v/v) were added. After mixing, the solution was incubated for 20 minutes.<sup>18-19</sup> The solution was measured the absorbance with a spectrophotometer at 412 nm. The standard used for the calibration curve was quercetin.<sup>20</sup> The total flavonoid content was expressed as micrograms of flavonoids per milligrams of the extract.

#### *Determination of the total tannin content*

A total of 30 mg of sample was weighed and put into a 10 mL volumetric flask. The solution of 0.5 ml was added with 3.0 ml of vanillin 4% and 1.5 ml of concentrated HCl. The mixture was incubated for 10 minutes.<sup>20-23</sup> The absorbance was measured with UV Vis spectrophotometer at 498 nm. The standard used for the calibration curve was catechin.<sup>24</sup> The total tannin content was expressed as micrograms of tannins per milligrams of the extract.

#### *Antioxidant activity test with DPPH method*

DPPH solution with 0.4 mM concentration was prepared, the absorbance was measured at 512 nm. The absorbance result of the DPPH solution was the absorbance control. The extract of sintok lancang leaves (sample) were first dissolved in methanol with five different concentrations of 10, 20, 30, 40, and 50 ppm. One ml of 0.4mM DPPH solution was placed into a 5 ml volumetric flask, then added with 4 ml of sample solution each of various concentrations. The mixture was stored for incubation for 30 min at room temperature, the absorbance was measured at maximum wavelength (512 nm).<sup>25-26</sup> The percent inhibition or DPPH scavenging effect was calculated using the following formula:

$$DPPH\ Scavenging\ effect\ \% = \frac{(A - B)}{A} \times 100\%$$

Where, A was the absorbance of DPPH solution and B was the absorbance of sample solution.

### *Antioxidant activity test with FRAP method*

Sample (0.2 g) dissolved with ethanol with 10 ml volumetric flask. Two ml of sample solution was added to 3 ml of FRAP reagent in a test tube, then incubation for 16 minutes. The absorbance was measured with UV Vis spectrophotometer at 595 nm. Antioxidant activity was expressed as  $\mu\text{mol trolox/g}$ .<sup>27,29</sup>

## **Results**

### *Phytochemical Qualitative Screening*

Phytochemical screening of *C. javanicum* leaves by using the following standard methods.<sup>13-16</sup> The results of the phytochemical qualitative test of sintok lancang leaves showed the presence of alkaloids using Mayer's reagents<sup>30</sup>, flavonoids using the Shinoda test<sup>31</sup>, tannins and steroids (Table 1.).

**Table 1.** The qualitative phytochemical of ethanolic extract *C. javanicum* leaves

<b>Phytochemical compound</b>	<b>Result</b>
Alkaloid	+
Flavonoid	+
Tannin	+
Steroid	+
Saponin	-

### *Antioxidant activity, total alkaloid, flavonoid and tannin content*

Ethanolic extract of *C. javanicum* leaves was conducted using percolation method. Antioxidant potential (DPPH & FRAP), total alkaloid content, total flavonoid content and total tannin content was calculated with standard protocols.

The results of antioxidant activity showed that the  $\text{IC}_{50}$  value of DPPH was  $26.99 \pm 0.27$  ppm quercetin equivalent while the FRAP method was  $779.73 \pm 19.66$   $\mu\text{mol trolox/g}$ . Total alkaloid,

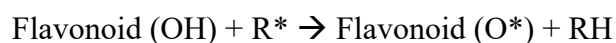
flavonoid, and tannin content respectively was  $20.82 \pm 1.31$   $\mu\text{g}$  caffeine equivalent/mg,  $76.62 \pm 1.22$   $\mu\text{g}$  quercetin equivalent/mg, and  $23.02 \pm 0.24$   $\mu\text{g}$  catechin equivalent/mg (Table 2.).

**Table 2.** Antioxidant activity, total alkaloids, flavonoids and tannins content of ethanolic extract *C. javanicum* leaves.

Sample	Ethanolic extract of <i>Cinnamomum javanicum</i>	Quercetin
Assay		
Antioxidant		
- DPPH (IC <sub>50</sub> ppm)	$26.99 \pm 0.27$	6.98
- FRAP ( $\mu\text{mol}$ trolox/g)	$779.73 \pm 19.66$	-
Total Alkaloid ( $\mu\text{g}$ caffeine equivalent/mg)	$20.82 \pm 1.31$	-
Total Flavonoid ( $\mu\text{g}$ quercetin equivalent/mg)	$76.62 \pm 1.22$	-
Total Tannin ( $\mu\text{g}$ catechin equivalent/mg)	$23.02 \pm 0.24$	-

## Discussion

Based on the results extract of *C. javanicum* leaves contain qualitative phytochemicals like flavonoid. Based on some literature flavonoids can prevent injury due to free radicals in various ways, one of which is the direct scavenging of free radicals. Flavonoids are oxidized by radicals, resulting in a more stable, less-reactive radical<sup>32-33</sup>, such as the following equation:



Besides that *C. javanicum* leaves contain alkaloid and tannin which may also potential contribute to the effectiveness of antioxidant.<sup>34-35</sup>

There hasn't been much study on *C. javanicum* leaves. However, some studies have stated that *C. javanicum* found in Sumatra can reduce fever, others claim that it has antioxidant potential but there are no studies of *C. javanicum* that grows in Kalimantan.<sup>36-37</sup>

According to some literature, antioxidant activity with DPPH method classified by IC<sub>50</sub> into: very strong (< 50 ppm), strong (50-100 ppm), moderate (101-150 ppm) and low (> 150 ppm)<sup>27,38</sup>, while antioxidant activity with FRAP method classified into very low FRAP (< 10



$\mu\text{mol/g}$ ), low FRAP (10-50  $\mu\text{mol/g}$ ), good FRAP (50-100  $\mu\text{mol/g}$ ), high FRAP (100-400  $\mu\text{mol/g}$ ), and very high FRAP ( $> 400 \mu\text{mol/g}$ ).<sup>39</sup> The ethanolic extract of *C. javanicum* leaves included in the very strong antioxidant activity ( $26.99 \pm 0.27 \text{ ppm}$ ) and very high FRAP ( $779.73 \pm 19.66 \mu\text{mol trolox/g}$ ). When compared with *C. javanicum* leaves study conducted in Malaysia with DPPH method (223.5 ppm)<sup>40</sup>, *C. javanicum* leaves in Indonesia, especially Central Kalimantan give better antioxidant activity.

Antioxidants are widely used for protection of various diseases such as coronary heart, cancer. Besides that, it is also often applied at the industrial level such as cosmetic for anti-aging and acne.<sup>41</sup> Based on the results *C. javanicum* leaves has the potential to be cultivated to keep sustainability and developed into antioxidant material.

## **Conclusion**

It can be concluded ethanolic extract of *C. javanicum* leaves has the potential high antioxidant activity tested by DPPH and FRAP methods. Furthermore, it is necessary to further identify the specific compounds that have antioxidant activity in *C. javanicum* leaves.

## **Acknowledgments**

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## **Conflict of interests**

The authors claim that there is no conflict of interest.

## **References**

1. Kumalasari A, Handayani W, Siswoyo TS, "Screening Fitokimia dan Studi Aktivitas Ekstrak Daun Sintok (*Cinnamomum sintoc* BI.) sebagai Antioksidan dan

- Antihiperlipidemia”, *Berkala Sainstek.*, VII (1): 24-27. (2019). doi: <https://doi.org/10.19184/bst.v7i1.9683>
2. Jiamworanunkul S, “Effective antioxidant production through submerged fermentation of edible mushrooms”, *Thai J Pharm Sci*, 43 (4), 213-218 (2019).
  3. Stagos D, “Antioxidant activity of polyphenolic plant extracts”, *Antioxidants* 9 (19) (2020). doi: <https://doi.org/10.3390/antiox9010019>
  4. Borquaye LS, Laryea MK, Gasu EN, Boateng MA, Baffour PK, Kyeremateng A, Doh G, “Anti-inflammatory and antioxidant activities of extracts of *Reissantia indica*, *Cissus cornifolia* and *Grosseria vignei*”, *Cogent Biology*, 6: 1785755 (2020). doi: <https://doi.org/10.1080/23312025.2020.1785755>
  5. Fajrin FA, Imandasari N, Barki T, Sulistyaningrum G, Afifah, Kristiningrum N, Puspitasari E, Holidah D, “The activity of red ginger oil in antioxidant study *in vitro* and antihyperalgesia effect in alloxan-induced painful diabetic neuropathy in mice”, *Thai J Pharm Sci*, 43 (2), 69-75 (2019).
  6. Leakaya N, Sato VH, Chewchinda S, “Antioxidant activity, total phenolic, total flavonoid content and HPTLC analysis of morin in *Maclura cochinchinensis* Heartwood extract”, *Thai J Pharm Sci*, 42 (Supplement Issue), 27-31 (2018).
  7. Al-Taie A, Victoria AO, “Supplementary medicines and antioxidants in viral infections: a review of proposed effects for COVID-19”, *Biomed Biotechnol Res J*, 4(Special Issue 1),19-24 (2020)
  8. Diniz LRL, Filho C da SMB, Fielding BC, Sousa DP de, “Natural Antioxidants: a review of studies oh human and animal coronavirus”, *Oxid Med Cell Longev*, 3173281 (2020). doi: <https://doi.org/10.1155/2020/3173281>

9. Gbaguidi AM, Chadare FJ, Salako VK, Idohou YO, Assogbadjo AE, “Optimisation of oven-drying of baobab leaves using a central composite design”, *Afr Crop Sci J*, 28 (Issue Supplement, S1), 15-26 (2020). doi: <https://doi.org/10.4314/acsj.v28i1.2S>
10. Mohamed Hanaa AR, Sallam YI, El-Leithy AS, Aly SE, “Lemongrass (*Cymbopogon citratus*) essential oil as affected by drying methods”, *Ann Agric Sci*, 57(2),113-116 (2020). doi: <http://dx.doi.org/10.1016/j.aogas.2012.08.004>
11. Thamkaew G, Sjöholm I, Galindo FG, “A review of drying methods for improving the quality of dried herbs”, *Crit Rev Food Sci Nutr*, (2020). doi: <https://doi.org/10.1080/10408398.2020.1765309>
12. Zhang QW, Lin LG, Ye WC, “Techniques for extraction and isolation of natural products: a comprehensive review”, *Chin Med*, 13:20 (2018). doi: <https://doi.org/10.1186/s13020-018-0177-x>
13. Vijayan K, Gopinathan M, Ambikapathy V, “Phytochemical screening and antioxidant activity of *Diospyros ebenum* J. koening ex retz, leaves extract”, *Int J Pharm Sci Res*, 11(10),5163-5169 (2020). doi: [https://doi.org/10.13040/IJPSR.0975-8232.11\(10\).5163-69](https://doi.org/10.13040/IJPSR.0975-8232.11(10).5163-69)
14. Ahmed Z, Aziz S, Hanif M, Mohiuddin SG, Khan SHA, Ahmed R, Ghadzi SMS, Bitar AN, “Phytochemical screening and enzymatic and antioxidant activities of *Erythrina suberosa* (Roxb) bark”, *J Pharm Bioallied Sci*, 12(2),192-200 (2020). doi: [https://doi.org/10.4103/jpbs.JPBS\\_222\\_19](https://doi.org/10.4103/jpbs.JPBS_222_19)
15. Ardhanay, SD, “Antibacterial activity of bawang dayak (*Eleutherine* sp.) and tawas ut (*Ampelocissus* sp.) from Central Kalimantan against *Propionibacterium acnes*”, *Int J App Pharm*, 11(Special issue 3),7-10 (2019). doi: <https://doi.org/10.22159/ijap.2019.v11s3.M0010>

16. Roghini R, Vijayalakshmi K, “Phytochemical screening, quantitative analysis of flavonoids and minerals in ethanolic extract of *Citrus paradise*”, *Int J Pharm Sci Res*, 9(11), 4859-4864 (2018). doi: [https://doi.org/10.13040/IJPSR.0975-8232.9\(11\).4859-64](https://doi.org/10.13040/IJPSR.0975-8232.9(11).4859-64)
17. Sathishkumar T, Baskar R, “Screening and quantification of phytochemicals in the leaves and flowers of *Tabernaemontana heyneana* Wall. A near threatened medicinal plant”, *Indian J Nat Prod Resour*, 5(3):237-243 (2014).
18. Sukmawati, Sudewi S, Pontoh J, “Optimasi dan validasi metode analisis dalam penentuan kandungan total flavonoid pada ekstrak daun geddi hijau (*Abelmoscus manihot* L.) yang diukur menggunakan spektrofotometri uv-vis”, *Pharmacon Jurnal Ilmiah Farmasi*, 7(3):32-41 (2018). doi: <https://doi.org/10.35799/pha.7.2018.20117>
19. Hasan AEZ, Nashrianto H, Juhaeni RN, Artika IM, “Optimization of conditions for flavonoids extraction from mangosteen (*Garcinia mangostana* L.)”, *Der Pharmacia Lettre*, 8(18),114-120 (2016).
20. Beyene BB, Alem FA, Ayana MT, “Determination of antioxidant and antibacterial activities of leaf extracts of *Plumbago zeylanica* (Amira)”, *Cogent Chemistry*, 6:1831715,1-16 (2020). doi: <https://doi.org/10.1080/23312009.2020.1831715>
21. Hayat J, Akodad M, Moumen A, Baghour M, Skalli A, Ezrari S, Belmalha S, “Phytochemical screening, polyphenols, flavonoids and tannin content, antioxidant activities and FTIR characterization of *Marrubium vulgare* L. from 2 different localities of Northeast of Morocco”, *Heliyon*, 6(11):e05609 (2020). doi: <https://doi.org/10.1016/j.heliyon.2020.e05609>
22. Rebaya A, Belghith SI, Baghdikian B, Leddet VM, Mabrouki F, Olivier E, Cherif JK, Ayadi MT, “Total phenolic, total flavonoid, tannin content, and antioxidant capacity of *Halimium halimifolium* (Cistaceae)”, *J App Pharm Sci*, 5(01),052-057 (2015). doi: <https://doi.org/10.7324/JAPS.2015.50110>

23. Medini F, Fellah H, Ksouri R, Abdelly C, “Total phenolic, flavonoid and tannin contents and antioxidant and antimicrobial activities of organic extracts of shoots of the plant *Limonium delicatulum*”, *J Taibah Univ Sci*, 8(2014),216-224 (2014).. doi: <https://doi.org/10.1016/j.jtusci.2014.01.003>
24. Formagio ASN, Volobuff CRF, Santiago M, Cardoso CAL, Vieira MDC, Pereira ZV, “Evaluation of antioxidant activity, total flavonoids, tanins and phenolic compounds in *Psychotria* leaf extracts”. *Antioxidants*,3: 745-757 (2014). doi: <https://doi.org/10.3390/antiox3040745>
25. Manssouri M, Znini M, Majidi L, “Studies on the antioxidant activity of essential oil and various extracts of *Ammodaucus leucotrichus* Coss. & Dur. fruits from Morocco”. *J Taibah Univ Sci*, 14(1),124-130 (2020). doi: <https://doi.org/10.1080/16583655.2019.1710394>
26. Ardhanay SD, Mulia DS, Rosawanti P, “Antioxidant activity of ethyl acetate fraction of *Macaranga triloba* leaves from Central Kalimantan”, *Asian J Pharm Clin Res*,11(Special issue 3),40-42 (2018). doi: <https://doi.org/10.22159/ajpcr.2018.v11s3.30026>
27. Sukweenadhi J, Yunita O, Setiawan F, Kartini, Siagian MT, Danduru AP, Avanti C, “Antioxidant activity screening of seven Indonesian herbal extract”, *Biodiversitas*,21(5),2062-2067 (2020). doi: <https://doi.org/10.13057/biodiv/d210532>
28. Andres AI, Petron MJ, Lopez AM, Timon ML, “Optimization of extraction conditions to improve phenolic content and in vitro antioxidant activity in craft brewers’ spent grain using response surface methodology (RSM)”, *Foods*, 9(1398), 1-13 (2020). doi: <https://doi.org/10.3390/foods9101398>
29. Mitrevska K, Grigorakis S, Loupassaki S, Calokerinos AC, “Antioxidant activity and polyphenolic content of North Macedonian wines”, *App sci*, 10:1-10 (2020). doi: <https://doi.org/10.3390/app10062010>

30. Senhaji S, Lamchouri F, Toufik H, “Phytochemical content, antibacterial and antioxidant potential of endemic plant *Anabasis aretioides* Coss. & Moq. (Chenopodiaceae)”, **Bio Res Int** 1-16 (2020). doi: <https://doi.org/10.1155/2020/6152932>
31. Mondal S, Rahaman ST, ”Flavonoids: a vital resource in healthcare and medicine”, **Pharm Pharmacol Int J**, 8(2),91-104 (2020). doi: <https://doi.org/10.15406/ppij.2020.08.00285>
32. Nijveldt RJ, Nood E van, Hoorn DEC van, Boelens PG, Norren K van, Leeuwen PAM van, “ Flavonoids: a review of probable mechanisms of action and potential applications”, **Am J Clin Nutr**,74, 418-25 (2001). doi: <https://doi.org/10.1093/ajcn/74.4.418>
33. Panche AN, Diwan AD, Chandra SR, “Flavonoids: an overview”, **J Nutr Sci**,5(e47):1-15 (2016). doi: <https://doi.org/10.1017/jns.2016.41>
34. Gan J, Feng Y, He Z, Li X, Zhang H, “Correlations between antioxidant activity and alkaloids and phenols of Maca (*Lepidium meyenii*)”, **J Food Quality**, Article ID 3185945, 1-10 (2017). doi: <https://doi.org/10.1155/2017/3185945>
35. Maisetta G, Batoni G, Caboni P, Esin S, Rinaldi AC, Zucca P, “Tannin profile, antioxidant properties, and antimicrobial activity of extracts from two Mediterranean species of parasitic plant *Cytinus*”, **BMC Comp & Alt Med**, 19(82),1-11 (2019). doi: <https://doi.org/10.1186/s12906-019-2487-7>
36. Ambri K, Afifuddin Y, Hafni A, “Exploration of medical plant in gunung leuser national park, sei betung resort, north sumatera”, **Peronema Forestry Science J**,4(2),19-32 (2015).
37. Kumar S, Kumari R, Mishra S, “Pharmacological properties and their medicinal uses of Cinnamomum: a review”, **J Pharm and Pharm**,71,1735-1761 (2019). doi: <https://doi.org/10.1111/jphp.13173>
38. Najafabadi SF, Safaeian L, Zolfaghari B, “In vitro antioxidant effects of different extracts obtained from the leaves and seeds of *Allium ampeloprasum* subsp. *persicum*”, **J Herb Pharm**,8(3), 256-260 (2019). doi: <https://doi.org/10.15171/jhp.2019.37>

39. Fernandes RPP, Trindade MA, Tonin FG, Lima CG, Pugine SMP, Munekata PES, Lorenzo JM, de Melo MP, “Evaluation of antioxidant capacity of 13 plant extracts by three different methods: cluster analyses applied for selection of the natural extracts with higher antioxidant capacity to replace synthetic antioxidant in lamb burgers”, *J Food Sci Technol*, 53(1), 451-460 (2016). doi: <https://doi.org/10.1007/s13197-015-1994-x>
40. Salleh WMN Hakimi Wan, Ahmad F, Yen KH, “Evaluation of antioxidant, anticholinesterase and antityrosinase activities of Malaysian *Cinnamomum* species”. *Dhaka Univ J Pharm Sci*,14(2),125-132 (2015).
41. Hassan A, Akmal Z, Khan N, “The phytochemical screening and antioxidants potential of *Schoenoplectus triqueter* L. Palla”, *J Chem*, 2020,1-8 (2020). doi: <https://doi.org/10.1155/2020/3865139>

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## THE PHYTOCHEMICAL SCREENING AND ANTIOXIDANT POTENTIAL OF *CINNAMOMUM JAVANICUM* BLUME LEAVES FROM CENTRAL KALIMANTAN

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**Background:** Sintok lancang (*Cinnamomum javanicum* Blume.) is one of the typical plant of Central Kalimantan, which has not been widely studied. Local people use leaves of *C. javanicum* to treat various diseases, like diabetes and skin diseases. Based on this, it is necessary to do preliminary study to know phytochemical content and antioxidant potential that may be contained in this plant. **Methods:** *C. javanicum* leaves were extracted using percolator with 96% ethanol. The extract was tested for qualitative phytochemical with standard procedures, while the antioxidant test was carried out using 1,1-diphenyl-2-picrylhydrazyl (DPPH) method with ascorbic acid as standard and ferric reducing antioxidant power (FRAP) with

### Introduction

Central Kalimantan is one of the tropical forest areas in Indonesia which has biodiversity especially natural medicine but has not been fully utilized. Traditional uses such as traditional medicine have been done, but further exploration and development have not been done much. Sintok lancang leaves (*Cinnamomum javanicum* Blume.) are one of the typical Central Kalimantan plants that have not been studied much, it's found in the Mungku Baru forest (Kawasan Hutan Dengan Tujuan Khusus) or educational forest managed by the Muhammadiyah University of Palangkaraya collaboration with the Borneo Nature Foundation (BNF). Local people use sintok lancang leaves for various diseases such as to treat abdomen pain, wounds, and diabetes.

There have not been many studies on *Cinnamomum javanicum*, especially those that grow in Indonesia, but one of the researches at the University of Jember states that different species, *Cinnamomum Sintoc* leaves has the potential as an antioxidant because it contains secondary metabolites such as flavonoids, alkaloids, steroids, terpenoids, and tannins which could prevent

the free radical.<sup>1-3</sup> Antioxidant use is associated with reduced production of ROS (Reactive oxygen species) and free radicals.<sup>4-6</sup> According to previous pieces of evidence, ROS have been implicated in the pathogenesis of various diseases such as cancer, aging, and other diseases. In addition, research related to natural antioxidants are considered to contribute to the therapeutic approach in the treatment of COVID-19, although their mechanism against COVID-19 is still unknown and speculative.<sup>7</sup> Several studies provide evidence that a reduction in ROS accumulation slows the apoptosis signaling activated by a coronavirus.<sup>8</sup>

In this study, the determination was carried out the total flavonoids, alkaloids and tannins compounds of *Cinnamomum javanicum* leaves. Antioxidant activity test was conducted by using DPPH and FRAP assay. The present data would help to explore the potential of Indonesia's natural plants, especially plants that have not been studied much so it can be further developed for the cosmetic, food and pharmaceutical industries.

## **Methods**

### *Plant Collection and Identification*

Fresh leaves of sintok lancang (Fig 1.) were collected in the Mungku Baru forest (Kawasan Hutan Dengan Tujuan Khusus) or educational forest managed by the Muhammadiyah University of Palangkaraya collaboration with the Borneo Nature Foundation (BNF), Palangka Raya, Central Kalimantan and authenticated by Dr. R. Hendrian, M.Sc from Indonesia Institute of Sciences, Research Center for Plant Conservation and Botanic Gardens, Bogor, Indonesia.



**Fig 1.** Sintok lancang (*Cinnamomum javanicum* Blume.)

#### *Preparation of Plant Extract*

The whole fresh leaves were washed, cleaned and dried in oven at 45<sup>0</sup>C.<sup>9-10</sup> Generally, the oven drying method is used for herb drying in the temperature range of 40-60<sup>0</sup>C.<sup>11</sup> The dried leaves were grinded and powdered. Extracted with ethanol 96% by percolation. Percolation is better than maceration because it is a continuous process in which the saturated solvent is constantly being replaced by fresh solvent.<sup>12</sup> The extract were concentrated in a rotary evaporator. The percentage yields (w/w) of the extracts were calculated using the formula:<sup>13</sup> (Weight of extract ÷ Weight of starting plant material) x 100%

#### *Phytochemical Qualitative Screening*

The ethanolic extract of sintok lancang leaves (*C. javanicum* Blume) was screened for potential presence of alkaloid, flavonoids, tannins, saponins, and steroids by using the following standard methods.<sup>14-17</sup>

#### *Determination of the total alkaloid content*

Ten mg extract of *C. javanicum* was weighed then dissolved in 10 ml ethanol. One ml extract was measured the absorbance with a spectrophotometer at 272 nm. The standard used for the calibration curve was caffeine.<sup>18</sup> The total alkaloid content was expressed as micrograms of alkaloids per milligrams of the extract.

#### *Determination of the total flavonoid content*

Five mg extract of *C. javanicum* was weighed then dissolved in 10 ml ethanol. One ml extract was placed in the volumetric flask, one ml AlCl<sub>3</sub> (2% b/v) and 8 ml acetic acid (5% v/v) were added. After mixing, the solution was incubated for 20 minutes.<sup>19-20</sup> The solution was measured the absorbance with a spectrophotometer at 412 nm. The standard used for the calibration curve was quercetin.<sup>21</sup> The total flavonoid content was expressed as micrograms of flavonoids per milligrams of the extract.

#### *Determination of the total tannin content*

A total of 30 mg of sample was weighed and put into a 10 ml volumetric flask. The solution of 0.5 ml was added with 3.0 ml of vanillin 4% and 1.5 ml of concentrated HCl. The mixture was incubated for 10 minutes.<sup>21-24</sup> The absorbance was measured with UV Vis spectrophotometer at 498 nm. The standard used for the calibration curve was catechin.<sup>25</sup> The total tannin content was expressed as micrograms of tannins per milligrams of the extract.

#### *Antioxidant activity test with DPPH method*

DPPH solution with 0.4 mM concentration was prepared, the absorbance was measured at 512 nm. The absorbance result of the DPPH solution was the absorbance control. The extract of sintok lancang leaves (sample) were first dissolved in methanol with five different concentrations of 10,20,30,40, and 50 ppm. One ml of 0.4mM DPPH solution was placed into a 5 ml volumetric flask, then added with 4 ml of sample solution each of various concentrations. The mixture was stored for incubation for 30 min at room temperature, the absorbance was measured at maximum wavelength (512 nm).<sup>26-27</sup> The percent inhibition or DPPH scavenging effect was calculated using the following formula:

$$DPPH\ Scavenging\ effect\ \% = \frac{(A - B)}{A} \times 100\%$$

Where, A was the absorbance of DPPH solution and B was the absorbance of sample solution.

28-29

#### *Antioxidant activity test with FRAP method*

Sample (0.2 g) dissolved with ethanol with 10 ml volumetric flask. Two ml of sample solution was added to 3 ml of FRAP reagent in a test tube, then incubation for 16 minutes. The absorbance was measured with UV Vis spectrophotometer at 595 nm. Antioxidant activity was expressed as  $\mu\text{mol trolox/g}$ .<sup>28,30</sup>

## **Results**

#### *Yield of The Extract*

Four hundred grams of *C. javanicum* fresh leaves were extracted into 51.1 g of extract. Based on the rendement calculation, the extraction of *C. javanicum* yields of 12.8%. Value yield is related to the number of secondary metabolites that successfully attracted when the extraction process<sup>13</sup>.

#### *Phytochemical Qualitative Screening*

Phytochemical screening of *C. javanicum* leaves by using the following standard methods.<sup>14-17</sup> The results of the phytochemical qualitative test of sintok lancang leaves showed the presence of alkaloids using Mayer's reagents<sup>31</sup>, flavonoids using the Shinoda test<sup>32</sup>, tannins and steroids (Table 1.).

**Table 1.** The qualitative phytochemical of ethanolic extract *C. javanicum* leaves

<b>Phytochemical compound</b>	<b>Result</b>
Alkaloids	+
Flavonoids	+
Tannins	+
Steroids	+
Saponins	-

### *Antioxidant activity, total alkaloid, flavonoid and tannin content*

Ethanollic extract of *C. javanicum* leaves was conducted using percolation method. Antioxidant potential (DPPH & FRAP), total alkaloid content, total flavonoid content and total tannin content was calculated with standard protocols.

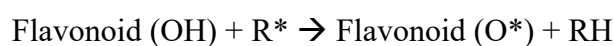
The results of antioxidant activity showed that the IC<sub>50</sub> value of DPPH was 26.99 ± 0.27 ppm quercetin equivalent while the FRAP method was 779.73 ± 19.66 µmol trolox/g. Total alkaloid, flavonoid, and tannin content respectively was 20.82 ± 1.31 µg caffeine equivalent/mg, 76.62 ± 1.22 µg quercetin equivalent/mg, and 23.02 ± 0.24 µg catechin equivalent/mg (Table 2.).

**Table 2.** Antioxidant activity, total alkaloids, flavonoids and tannins content of ethanolic extract *C. javanicum* leaves.

Sample	Ethanollic extract of <i>Cinnamomum javanicum</i>	Quercetin
Assay		
Antioxidant		
- DPPH (IC <sub>50</sub> ppm)	26.99 ± 0.27	6.98
- FRAP (µmol trolox/g)	779.73 ± 19.66	-
Total Alkaloid (µg caffeine equivalent/mg)	20.82 ± 1.31	-
Total Flavonoid (µg quercetin equivalent/mg)	76.62 ± 1.22	-
Total Tannin (µg catechin equivalent/mg)	23.02 ± 0.24	-

### **Discussion**

Based on the results extract of *C. javanicum* leaves contain qualitative phytochemicals like flavonoid. Based on some literature flavonoids can prevent injury due to free radicals in various ways, one of which is the direct scavenging of free radicals. Flavonoids are oxidized by radicals, resulting in a more stable, less-reactive radical<sup>33-34</sup>, such as the following equation:



Besides that *C. javanicum* leaves contain alkaloids and tannins which may also potential contribute to the effectiveness of antioxidant.<sup>35-36</sup> Alkaloids have antioxidant properties

through capturing free radicals, or binding to catalysts involved indifferent oxidation processes occurring within the human body for preventing a variety of degenerative diseases.<sup>37</sup> The free radical scavenging activity of tannins showed by the ability of antioxidant to donate electron to a free radical and produce a more stable and therefore less harmful radical structure. Tannins are able to bond cations of transition metals and act as protective agents against progression of some diseases, e.g. Alzheimer's or Parkinson's disease. Antioxidant activity of tannins can also be exhibited through inhibition of prooxidative enzymes.<sup>38</sup>

There hasn't been much study on *C. javanicum* leaves. However, some studies have stated that *C. javanicum* found in Sumatra can reduce fever, others claim that it has antioxidant potential but there are no studies of *C. javanicum* that grows in Kalimantan.<sup>39-40</sup>

Yuan et al stated that *C. javanicum* both leaf and stem extract showed antimicrobial activity against *Listeria monocytogenes*.<sup>41</sup> The essential oil of *C. javanicum* also showed antibacterial activities against four strains of food pathogenic bacteria: *Staphylococcus aureus* (ATCC 29213), *Listeria monocytogenes* (ATCC 7644), *Salmonella typhimurium* (ATCC 25922) and *Salmonella enteritidis* (ATCC 29213).<sup>42</sup> The various study results can be the basis for further research.

According to some literature, antioxidant activity with DPPH method classified by IC<sub>50</sub> into: very strong (< 50 ppm), strong (50-100 ppm), moderate (101-150 ppm) and low (> 150 ppm)<sup>28,43</sup>, while antioxidant activity with FRAP method classified into very low FRAP (< 10 µmol/g), low FRAP (10-50 µmol/g), good FRAP (50-100 µmol/g), high FRAP (100-400 µmol/g), and very high FRAP (> 400 µmol/g).<sup>44</sup> The ethanolic extract of *C. javanicum* leaves included in the very strong antioxidant activity (26.99 ± 0.27 ppm) and very high FRAP (779.73 ± 19.66 µmol trolox/g). When compared with *C. javanicum* leaves study conducted in Malaysia with DPPH method (223.5 ppm)<sup>45</sup>, *C. javanicum* leaves in Indonesia, especially Central Kalimantan give better antioxidant activity.



DPPH is a stable free radical with an unpaired electron that is delocalized over the entire molecule. The DPPH assay is based on both electron transfer and hydrogen atom transfer reactions. The benefits of the DPPH assay is easy to do, rapid method and economic. Even though the DPPH assay is simple, its sensitivity may be affected by several factors, such as the type of solvent, reaction time, temperature and freshness of DPPH reagent,<sup>46,47</sup> while the FRAP assay is a non-specific, redox-linked, colorimetric assay that is related to the molar concentration of the antioxidant present. The FRAP assay is a typical electron transfer based method that measures the reduction of ferric ion ( $\text{Fe}^{3+}$ )-ligand complex to the ferrous ( $\text{Fe}^{2+}$ ) complex by antioxidants in acidic media. One limitation of FRAP assay is the tendency to precipitate, forming a suspension and staining the measuring cuvette. Therefore, time to add  $\text{FeCl}_3$  is essential to prevent error interpretation. However, the FRAP or DPPH assay is simple, economic, rapid and not require specialized equipment.<sup>47</sup>

Antioxidants are widely used for protection of various diseases such as coronary heart, cancer. Besides that, it is also often applied at the industrial level such as cosmetic for anti-aging and acne.<sup>48</sup> The role of the food industry and mass media were explored to focus on health promotion of the older person in the present.<sup>49</sup>

Based on the results *C. javanicum* leaves has the potential to be cultivated to keep sustainability and developed into antioxidant material.

## **Conclusion**

It can be concluded ethanolic extract of *C. javanicum* leaves has the potential high antioxidant activity tested by DPPH and FRAP methods. Furthermore, it is necessary to further identify the specific compounds that have antioxidant activity in *C. javanicum* leaves.

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### Conflict of interests

The authors claim that there is no conflict of interest.

### References

1. Kumalasari A, Handayani W, Siswoyo TS, “Screening Fitokimia dan Studi Aktivitas Ekstrak Daun Sintok (*Cinnamomum sintoc* BI.) sebagai Antioksidan dan Antihiperlipidemia”, *Berkala Sainstek*, VII(1), 24-27 (2019). doi: <https://doi.org/10.19184/bst.v7i1.9683>
2. Jiamworanunkul S, “Effective antioxidant production through submerged fermentation of edible mushrooms”, *Thai J Pharm Sci*, 43(4), 213-218 (2019).
3. Stagos D, “Antioxidant activity of polyphenolic plant extracts”, *Antioxidants*, 9(19) (2020). doi: <https://doi.org/10.3390/antiox9010019>
4. Borquaye LS, Laryea MK, Gasu EN, Boateng MA, Baffour PK, Kyeremateng A, Doh G, “Anti-inflammatory and antioxidant activities of extracts of *Reissantia indica*, *Cissus cornifolia* and *Grosseria vignei*”, *Cogent Biology*, 6: 1785755 (2020). doi: <https://doi.org/10.1080/23312025.2020.1785755>
5. Fajrin FA, Imandasari N, Barki T, Sulistyaningrum G, Afifah, Kristiningrum N, Puspitasari E, Holiday D, “The activity of red ginger oil in antioxidant study *in vitro* and antihyperalgesia effect in alloxan-induced painful diabetic neuropathy in mice”, *Thai J Pharm Sci*, 43(2), 69-75 (2019).

6. Leakaya N, Sato VH, Chewchinda S, “Antioxidant activity, total phenolic, total flavonoid content and HPTLC analysis of morin in *Maclura cochinchinensis* Heartwood extract”, *Thai J Pharm Sci*, 42(Supplement Issue), 27-31 (2018).
7. Al-Taie A, Victoria AO, “Supplementary medicines and antioxidants in viral infections: a review of proposed effects for COVID-19”, *Biomed Biotechnol Res J*, 4(Special Issue 1), 19-24 (2020)
8. Diniz LRL, Filho C da SMB, Fielding BC, Sousa DP de, “Natural Antioxidants: a review of studies oh human and animal coronavirus”, *Oxid Med Cell Longev*, 3173281 (2020). doi: <https://doi.org/10.1155/2020/3173281>
9. Gbaguidi AM, Chadare FJ, Salako VK, Idohou YO, Assogbadjo AE, “Optimisation of oven-drying of baobab leaves using a central composite design”, *Afc Crop Sci J*, 28(Issue Supplement, S1), 15-26 (2020). doi: <https://doi.org/10.4314/acsj.v28i1.2S>
10. Mohamed Hanaa AR, Sallam YI, El-Leithy AS, Aly SE, “Lemongrass (*Cymbopogon citratus*) essential oil as affected by drying methods”, *Ann Agric Sci*, 57(2), 113-116 (2020). doi: <http://dx.doi.org/10.1016/j.aosas.2012.08.004>
11. Thamkaew G, Sjöholm I, Galindo FG, “A review of drying methods for improving the quality of dried herbs”, *Crit Rev Food Sci Nutr*, (2020). doi: <https://doi.org/10.1080/10408398.2020.1765309>
12. Zhang QW, Lin LG, Ye WC, “Techniques for extraction and isolation of natural products: a comprehensive review”, *Chin Med*, 13:20 (2018). doi: <https://doi.org/10.1186/s13020-018-0177-x>
13. Kusuma SAF, Mita SR, Firdayani I, Mustarichie R, “Study on the antibacterial activity of fruit extracts of klutuk banana (*Musa balbisiana* Colla) against *Shigella dysenteriae* ATCC 13313”, *Asian J Pharm Clin Res*, 10(7), 220-223 (2017). doi: <https://doi.org/10.22159/ajpcr.2017.v10i7.18561>

14. Vijayan K, Gopinathan M, Ambikapathy V, “Phytochemical screening and antioxidant activity of *Diospyros ebenum* J. koening ex retz, leaves extract”, *Int J Pharm Sci Res*, 11(10), 5163-5169 (2020). doi: [https://doi.org/10.13040/IJPSR.0975-8232.11\(10\).5163-69](https://doi.org/10.13040/IJPSR.0975-8232.11(10).5163-69)
15. Ahmed Z, Aziz S, Hanif M, Mohiuddin SG, Khan SHA, Ahmed R, Ghadzi SMS, Bitar AN, “Phytochemical screening and enzymatic and antioxidant activities of *Erythrina suberosa* (Roxb) bark”, *J Pharm Bioallied Sci*, 12(2), 192-200 (2020). doi: [https://doi.org/10.4103/jpbs.JPBS\\_222\\_19](https://doi.org/10.4103/jpbs.JPBS_222_19)
16. Ardhanay, SD, “Antibacterial activity of bawang dayak (*Eleutherine* sp.) and tawas ut (*Ampelocissus* sp.) from Central Kalimantan against *Propionibacterium acnes*”, *Int J App Pharm*, 11(Special issue 3), 7-10 (2019). doi: <https://doi.org/10.22159/ijap.2019.v11s3.M0010>
17. Roghini R, Vijayalakshmi K, “Phytochemical screening, quantitative analysis of flavonoids and minerals in ethanolic extract of *Citrus paradise*”, *Int J Pharm Sci Res*, 9(11), 4859-4864 (2018). doi: [https://doi.org/10.13040/IJPSR.0975-8232.9\(11\).4859-64](https://doi.org/10.13040/IJPSR.0975-8232.9(11).4859-64)
18. Sathishkumar T, Baskar R, “Screening and quantification of phytochemicals in the leaves and flowers of *Tabernaemontana heyneana* Wall. A near threatened medicinal plant”, *Indian J Nat Prod Resour*, 5(3), 237-243 (2014).
19. Sukmawati, Sudewi S, Pontoh J, “Optimasi dan validasi metode analisis dalam penentuan kandungan total flavonoid pada ekstrak daun geddi hijau (*Abelmoscus manihot* L.) yang diukur menggunakan spektrofotometri uv-vis”, *Pharmacon Jurnal Ilmiah Farmasi*, 7(3), 32-41 (2018). doi: <https://doi.org/10.35799/pha.7.2018.20117>
20. Hasan AEZ, Nashrianto H, Juhaeni RN, Artika IM, “Optimization of conditions for flavonoids extraction from mangosteen (*Garcinia mangostana* L.)”, *Der Pharmacia Lettre*, 8(18), 114-120 (2016).

21. Beyene BB, Alem FA, Ayana MT, “Determination of antioxidant and antibacterial activities of leaf extracts of *Plumbago zeylanica* (Amira)”, **Cogent Chemistry**, 6:1831715,1-16 (2020). doi: <https://doi.org/10.1080/23312009.2020.1831715>
22. Hayat J, Akodad M, Moumen A, Baghour M, Skalli A, Ezrari S, Belmalha S, “Phytochemical screening, polyphenols, flavonoids and tannin content, antioxidant activities and FTIR characterization of *Marrubium vulgare* L. from 2 different localities of Northeast of Morocco”, **Heliyon**, 6(11), e05609 (2020). doi: <https://doi.org/10.1016/j.heliyon.2020.e05609>
23. Rebaya A, Belghith SI, Baghdikian B, Leddet VM, Mabrouki F, Olivier E, Cherif JK, Ayadi MT, “Total phenolic, total flavonoid, tannin content, and antioxidant capacity of *Halimium halimifolium* (Cistaceae)”, **J App Pharm Sci**, 5(01), 052-057 (2015). doi: <https://doi.org/10.7324/JAPS.2015.50110>
24. Medini F, Fellah H, Ksouri R, Abdelly C, “Total phenolic, flavonoid and tannin contents and antioxidant and antimicrobial activities of organic extracts of shoots of the plant *Limonium delicatulum*”, **J Taibah Univ Sci**, 8, 216-224 (2014). doi: <https://doi.org/10.1016/j.jtusci.2014.01.003>
25. Formagio ASN, Volobuff CRF, Santiago M, Cardoso CAL, Vieira MDC, Pereira ZV, “Evaluation of antioxidant activity, total flavonoids, tanins and phenolic compounds in *Psychotria* leaf extracts”. **Antioxidants**, 3, 745-757 (2014). doi: <https://doi.org/10.3390/antiox3040745>
26. Manssouri M, Znini M, Majidi L, “Studies on the antioxidant activity of essential oil and various extracts of *Ammodaucus leucotrichus* Coss. & Dur. fruits from Morocco”. **J Taibah Univ Sci**, 14(1), 124-130 (2020). doi: <https://doi.org/10.1080/16583655.2019.1710394>

27. Ardhany SD, Mulia DS, Rosawanti P, “Antioxidant activity of ethyl acetate fraction of *Macaranga triloba* leaves from Central Kalimantan”, *Asian J Pharm Clin Res*, 11(Special issue 3), 40-42 (2018). doi: <https://doi.org/10.22159/ajpcr.2018.v11s3.30026>
28. Sukweenadhi J, Yunita O, Setiawan F, Kartini, Siagian MT, Danduru AP, Avanti C, “Antioxidant activity screening of seven Indonesian herbal extract”, *Biodiversitas*, 21(5), 2062-2067 (2020). doi: <https://doi.org/10.13057/biodiv/d210532>
29. Andres AI, Petron MJ, Lopez AM, Timon ML, “Optimization of extraction conditions to improve phenolic content and in vitro antioxidant activity in craft brewers’ spent grain using response surface methodology (RSM)”, *Foods*, 9(1398), 1-13 (2020). doi: <https://doi.org/10.3390/foods9101398>
30. Mitrevska K, Grigorakis S, Loupassaki S, Calokerinos AC, “Antioxidant activity and polyphenolic content of North Macedonian wines”, *App sci*, 10, 1-10 (2020). doi: <https://doi.org/10.3390/app10062010>
31. Senhaji S, Lamchouri F, Toufik H, “Phytochemical content, antibacterial and antioxidant potential of endemic plant *Anabasis aretioides* Coss. & Moq. (Chenopodiaceae)”, *Bio Res Int*, 1-16 (2020). doi: <https://doi.org/10.1155/2020/6152932>
32. Mondal S, Rahaman ST, ”Flavonoids: a vital resource in healthcare and medicine”, *Pharm Pharmacol Int J*, 8(2), 91-104 (2020). doi: <https://doi.org/10.15406/ppij.2020.08.00285>
33. Nijveldt RJ, Nood E van, Hoorn DEC van, Boelens PG, Norren K van, Leeuwen PAM van, “Flavonoids: a review of probable mechanisms of action and potential applications”, *Am J Clin Nutr*, 74, 418-25 (2001). doi: <https://doi.org/10.1093/ajcn/74.4.418>
34. Panche AN, Diwan AD, Chandra SR, “Flavonoids: an overview”, *J Nutr Sci*, 5(e47), 1-15 (2016). doi: <https://doi.org/10.1017/jns.2016.41>

35. Gan J, Feng Y, He Z, Li X, Zhang H, “Correlations between antioxidant activity and alkaloids and phenols of Maca (*Lepidium meyenii*)”, *J Food Quality*, Article ID 3185945, 1-10 (2017). doi: <https://doi.org/10.1155/2017/3185945>
36. Maisetta G, Batoni G, Caboni P, Esin S, Rinaldi AC, Zucca P, “Tannin profile, antioxidant properties, and antimicrobial activity of extracts from two Mediterranean species of parasitic plant *Cytinus*”, *BMC Comp & Alt Med*, 19(82), 1-11 (2019). doi: <https://doi.org/10.1186/s12906-019-2487-7>
37. Thawabteh A, Juma S, Bader M, Karaman D, Scrano L, Bufo AS, Karaman R, “The biological activity of natural alkaloids against herbivores, cancerous cells and pathogens”, *Toxins*, 11(656), 1-28 (2019). doi: <https://doi.org/10.3390/toxins11110656>
38. Macáková K, Kolečkář V, Cahlíková L, Chlebek J, Hošťálková A, Kuča K, Jun D, Opletal L, “Tannins and their influence on health” *Recent Adv Medicinal Chemist*, 1, 159-208 (2014). doi: <https://doi.org/10.1016/B978-0-12-803961-8.50006-3>
39. Ambri K, Afifuddin Y, Hafni A, “Exploration of medical plant in gunung leuser national park, sei betung resort, north sumatera”, *Peronema Forestry Science J*, 4(2), 19-32 (2015).
40. Kumar S, Kumari R, Mishra S, “Pharmacological properties and their medicinal uses of Cinnamomum: a review”, *J Pharm and Pharm*, 71, 1735-1761 (2019). doi: <https://doi.org/10.1111/jphp.13173>
41. Yuan W, Lee HW, Yuk HG, “Antimicrobial efficacy of *Cinnamomum javanicum* plant extract against *Listeria monocytogenes* and its application potential with smoked salmon”, *Int J Food Microbiology*, 260, 42-50 (2017). doi: <https://doi.org/10.1016/j.ijfoodmicro.2017.08.015>

42. Vairappan CS, Nagappan T, Kulip J, “The essential oils profiles and antibacterial activity of six wild *Cinnamomum* species”, *Nat Prod Commun*, 9(9), 1387-1389 (2014). doi: <https://doi.org/10.1177/1934578X1400900941>
43. Najafabadi SF, Safaeian L, Zolfaghari B, “In vitro antioxidant effects of different extracts obtained from the leaves and seeds of *Allium ampeloprasum* subsp. *persicum*”, *J Herb Pharm*,8(3), 256-260 (2019). doi: <https://doi.org/10.15171/jhp.2019.37>
44. Fernandes RPP, Trindade MA, Tonin FG, Lima CG, Pugine SMP, Munekata PES, Lorenzo JM, de Melo MP, “Evaluation of antioxidant capacity of 13 plant extracts by three different methods: cluster analyses applied for selection of the natural extracts with higher antioxidant capacity to replace synthetic antioxidant in lamb burgers”, *J Food Sci Technol*, 53(1), 451-460 (2016). doi: <https://doi.org/10.1007/s13197-015-1994-x>
45. Salleh WMN, Hakimi Wan, Ahmad F, Yen KH, “Evaluation of antioxidant, anticholinesterase and antityrosinase activities of Malaysian *Cinnamomum* species”. *Dhaka Univ J Pharm Sci*,14(2),125-132 (2015).
46. Liang N, Kitts DD, “Antioxidant property of coffee components: assessment of methods that define mechanisms of action”, *Molecules*, 19, 19180-19208 (2014). doi: <https://doi.org/10.3390/molecules191119180>
47. Shahidi F, Zhong Y, “Measurement of antioxidant activity”, *J Functional Foods* , 18(Part B), 757-781 (2015). doi: <https://doi.org/10.1016/j.jff.2015.01.047>
48. Hassan A, Akmal Z, Khan N, “The phytochemical screening and antioxidants potential of *Schoenoplectus triqueter* L. Palla”, *J Chem*, 2020,1-8 (2020). doi: <https://doi.org/10.1155/2020/3865139>
49. Wilson DW, Nash P, Buttar HS, Griffiths K, Singh R, Meester FD, Horiuchi R, Takahashi T, “The role of food antioxidants, benefits of functional foods, and influence of feeding



habits on the health of the older person: an overview”, *Antioxidants*, 6 (4): 81 (2017). doi:

<https://doi.org/10.3390/antiox6040081>



**UNIVERSITAS SARI MULIA**  
**LEMBAGA PENELITIAN DAN PENGABDIAN KEPADA MASYARAKAT**  
**DEWAN KOMITE ETIK PENELITIAN**

Jl. Pramuka No.02 Banjarmasin Tlp. (0511) 3268105

Banjarmasin, 21 Juli 2020

No. SK : 072/KE-LPPM/UNISM/VII/2020  
Lampiran : -  
Perihal : Rekomendasi Penelitian

Peneliti yang disebutkan dibawah ini :

Ketua Peneliti : Syahrida Dian Ardhany  
NIP/NIK/NIM : 14.0601.033  
Anggota Peneliti : 1. Susi Novaryatiin  
2. Nanang Hanafi

Judul Penelitian : Skrining fitokimia dan potensi antioksidan daun sintok  
(*Cinnamomum javanicum* Blume) asal Kalimantan Tengah

Berdasarkan pertimbangan Dewan Komite Etik Penelitian diputuskan bahwa Peneliti yang disebutkan diatas telah **DISETUJUI** untuk melanjutkan penelitiannya.

Demikian surat persetujuan ini diterbitkan untuk dipergunakan dengan penuh tanggung jawab.

Menyetujui,

An. Ketua

Sekretaris Dewan Komite Etik Penelitian



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